



Multiple recent co-options of Optix associated with novel traits in adaptive butterfly wing radiations

Martin *et al.*

RESEARCH

Open Access

Multiple recent co-options of Optix associated with novel traits in adaptive butterfly wing radiations

Arnaud Martin^{1,2*}, Kyle J McCulloch², Nipam H Patel³, Adriana D Briscoe², Lawrence E Gilbert⁴ and Robert D Reed¹

Abstract

Background: While the ecological factors that drive phenotypic radiations are often well understood, less is known about the generative mechanisms that cause the emergence and subsequent diversification of novel features. *Heliconius* butterflies display an extraordinary diversity of wing patterns due in part to mimicry and sexual selection. Identifying the genetic drivers of this crucible of evolution is now within reach, as it was recently shown that cis-regulatory variation of the *optix* transcription factor explains red pattern differences in the adaptive radiations of the *Heliconius melpomene* and *Heliconius erato* species groups.

Results: Here, we compare the developmental expression of the Optix protein across a large phylogenetic sample of butterflies and infer that its color patterning role originated at the base of the neotropical passion-vine butterfly clade (Lepidoptera, Nymphalidae, Tribe: Heliconiini), shortly predating multiple Optix-driven wing pattern radiations in the speciose *Heliconius* and *Eueides* genera. We also characterize novel Optix and Doublesex expression in the male-specific pheromone wing scales of the basal heliconiines *Dryas* and *Agraulis*, thus illustrating that within the Heliconiini lineage, Optix has been evolutionarily redeployed in multiple contexts in association with diverse wing features.

Conclusions: Our findings reveal that the repeated co-option of Optix into various aspects of wing scale specification was associated with multiple evolutionary novelties over a relatively short evolutionary time scale. In particular, the recruitment of Optix expression in colored scale cell precursors was a necessary condition to the explosive diversification of passion-vine butterfly wing patterns. The novel deployment of a gene followed by spatial modulation of its expression in a given cell type could be a common mode of developmental innovation for triggering phenotypic radiations.

Keywords: *Heliconius*, Adaptive radiation, Evolutionary novelty, Pattern evolution, Co-option, Sexual dimorphism

Background

Adaptive or phenotypic radiations are defined by ‘the evolution of phenotypic diversity within a rapidly multiplying lineage’ [1,2] and form crucibles of natural variation for the study of distinctness, as illustrated by Darwin’s work on Galapagos finches [3]. Ecological arguments involving external (biotic or abiotic) factors are traditionally invoked to explain why phenotypic disparity is maintained over time [2], but they fail to explain the

variation-generating mechanisms that allow these rapid bouts of evolution to occur. Recently, a handful of studies focusing on the *intrinsic causes* of adaptive radiations showed that diversification could follow relatively simple developmental innovations [4-9]. What principles underlie the emergence of new features, and how do these features further diversify [10,11]? A meaningful answer to this question may come from comparisons between many organisms, and adaptive radiations assist us in this task by providing a replicated and diverse template of phenotypic variation [12]. *Heliconius* butterflies are perhaps one of the best studied examples of a phenotypic radiation [13-15]. The genus comprises 40 species distributed across the Neotropics and is characterized by a variegated mosaic of wing patterns that vary both among

* Correspondence: heliconiuswing@gmail.com

¹Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853, USA

²Department of Ecology and Evolutionary Biology, University of California, Irvine, CA 92697-2525, USA

Full list of author information is available at the end of the article

and within species [13,16]. Wing patterns commonly function in nature as aposomatic traits (that is, as signals displayed to warn potential predators of danger), and butterflies such as *Heliconius erato* and *Heliconius melpomene* are a well-known example of this phenomenon [17]. Both species signal their unpalatability by displaying similar wing patterns that are indistinguishable from each other, are endemic to a restricted geographical range, and almost always include red coloration [18]. This Müllerian (that is, reciprocal) mimicry provides a strong adaptive advantage and predators learn to avoid all co-mimics [19]. In fact, most *Heliconius* species display extensive geographic variation in discrete wing patterns; more than 300 *Heliconius* regional morphs are recognized by their distinct wing pattern phenotypes and are each locally involved in complex mimicry rings with other co-mimetic butterflies [16,19,20].

Many ecological factors explain the diversity of *Heliconius* butterflies [21] such as their co-evolution with host plants of the *Passiflora* genus [22,23] and their participation in dynamic co-mimetic communities [19,24]. However, these extrinsic factors remain mute regarding the mechanisms that generate wing pattern variation in the first place. Mapping experiments taking advantage of intraspecific variability in the genus have begun to shed light on this question, and have revealed a monogenic (Mendelian) genetic architecture for discrete allelic variation in red patterns [25-27]. In particular, it was found that cis-regulatory evolution of the *optix* transcription factor is responsible for the presence/absence of red wing patterns in convergent forms of the *H. erato* and *H. melpomene* species, as well as in the divergent *Heliconius cydno* and *Heliconius pacheus* morphs [28,29]. Genetic variation at the *optix* locus explains a large fraction of the diversity of wing patterns in the genus, and was also found to contribute to the convergent evolution of mimetic forms by introgression of *optix* alleles in related lineages, resulting in a scattered phylogenetic distribution of adaptive patterns [15,18,30-32]. Thus, *optix* incarnates an emblematic example of 'hotspot of evolution' where the same gene has repeatedly caused phenotypic variation [28,33]. Interestingly, some of the variable red patterns controlled by *optix* are not only involved in predation avoidance, but are also drivers of reproductive isolation by assortative mating [34-36], suggesting that *optix* allelic variation may directly influence speciation rates.

Despite their ecological significance, little is known about the evolutionary origins of *optix*-positive red color patterns. Drawing conclusions from morphological analyses alone has been challenging because the wing pattern architecture of *Heliconius* is divergent relative to the nymphalid ground plan [27,37,38] - the system of wing pattern homologies that is otherwise broadly

conserved across the family Nymphalidae [39,40]. Here, we generated a polyclonal antibody directed against the *Optix* protein and compared its immunoreactivity in the developing wings of *Heliconius* and other butterflies. We found that the role of *Optix* in color patterning is relatively recent, making it a synapomorphy of neotropical passion-vine butterflies (tribe: Heliconiini), and that *Optix* was also associated to the development of novel, sexually dimorphic scale structures in basal heliconiines. In the light of previous work, we conclude that co-option of the *Optix* transcription factor into the color pattern formation process was a key pre-requisite for the diversification of red wing patterns. The co-option of *Optix* is thus consistent with a two-step innovation/diversification model, where the evolution of a novel character followed by its spatial modulation generated a wide array of distinct wing patterns.

Methods

Polyclonal antibody production

The nucleotide sequence of the *H. erato optix* gene [GenBank: JN102349.1] was modified using codon degeneracy to artificially lower high GC content without affecting the native amino-acid sequence. The corresponding DNA fragment was generated by gene synthesis (Genewiz, South Plainfield, New Jersey, USA) and cloned into the pQE-30 expression vector (Qiagen, Valencia, California, USA). A 6xHis-*Optix* fusion protein was expressed in bacterial cultures, extracted in 8 M Urea and purified using Ni²⁺ affinity column. An insoluble precipitate was recovered upon dialysis and approximately 1 mg was injected in a rat for immunization (5 injections of 200 ug each across 69 days; Covance, Princeton, New Jersey, USA).

Butterfly tissues

Butterfly pupae either originated from individuals collected in southern California, from laboratory colonies maintained at University of Texas - Austin, or from a butterfly farm in Costa Rica [see Additional file 1: Table S1]. Whenever possible, the time and day of pupation was monitored. Pupae were then kept at room temperature (23 to 26°C) and dissected on the fourth or fifth day after pupation, a stage at which wings are resistant to shearing while also being permeable to antibodies. Individuals originating from Costa Rica were shipped as untimed pupae and arrived in the laboratory after three days of transit. These pupae were approximately staged by screening their external eye morphology upon receipt. Pupae not presenting mature ommatidial structures (that is, those at an earlier stage of development) were immediately dissected. A portion of these then yielded pupal wing tissues at stages that were early enough for antibody permeability [see Additional file 1: Table S1]. The immunohistochemical

signal gradually decreases as pupal development proceeds to a point where scale cells begin to mature, the wing epithelia become an opaque white, and the forewings easily detach from the cuticle upon dissection. For all species, dissections performed before this stage resulted in reproducible immunodetection of Optix.

Tissue preparation and immunohistochemistry

Pupae were cold-anesthetized and dissected in PBS. Pupal forewings were incubated in fixative (PBS, ethylene glycol 5 mM tetraacetic acid, 0.01% Triton X-100, 1.85% formaldehyde) for 30 min at room temperature while still attached to their cuticle. Similarly, and in order to maintain flat wing tissues, hindwings were kept attached to their pupal case and fixed for at least 5 min as such, detached by dissection, and then the remainder of the 30 min fixation was completed on free-floating wings. Fixed wings were washed five times with PBST (PBS, 0.5% Triton X-100) and stored at 4°C for up to two months. Wings were cleaned from remnants of peripodial membrane with fine forceps and incubated in blocking buffer for 2 hrs (PBS, 0.5% Triton X-100, 5% Normal Goat Serum; at room temperature), incubated overnight at 4°C with anti-Optix polyclonal rat serum (dilution 1:3000), washed six times with PBST, incubated for 2 hrs at room temperature with an anti-rat IgG Alexa555 (Cell Signaling Technology, Danvers, Massachusetts, USA; 1:2000 dilution), and washed extensively in PBST. Nuclear counterstaining was performed with DAPI, and the wings were mounted in glycerol (PBS, 60% glycerol, 2 mM ethylenediaminetetraacetic acid). F-actin counterstaining was performed using phalloidin conjugated to green fluorescent Oregon Green 488 dye (Life Technologies, Carlsbad, California, USA; 1:400 dilution). For immunostaining in eyes and optic lobes, whole heads were fixed in 4% paraformaldehyde, sucrose protected, and frozen for sectioning. Horizontal sections of 14- μ m thickness were made using a cryostat and used for immunohistofluorescence using a previously described procedure [41]. A rabbit anti-UV opsin primary antibody [41] and an anti-rabbit IgG Alexa488 secondary antibody (Cell Signaling Technology, Danvers, Massachusetts, USA; 1:2000 dilution) were used as a positive control in adult eyes. The anti-Doublesex monoclonal mouse antibody [42] was a kind gift of C. Robinett (Janelia Farms Research Campus), was used at a 1:200 dilution and was detected using an anti-mouse IgG Alexa 488 secondary antibody (Cell Signaling Technology, Danvers, Massachusetts, USA; 1:2000 dilution).

Microscopy

Stained tissues were imaged with a Zeiss Axioplan 2 epifluorescence microscope and with a Zeiss LSM 700

confocal microscope. Tiled images were assembled using either the Photomerge function of Adobe Photoshop CS3 (epifluorescence imaging), or were acquired using the Tiled Scan function in the Zeiss ZEN software (confocal imaging). Images were submitted to a linear contrast adjustment to eliminate background illumination.

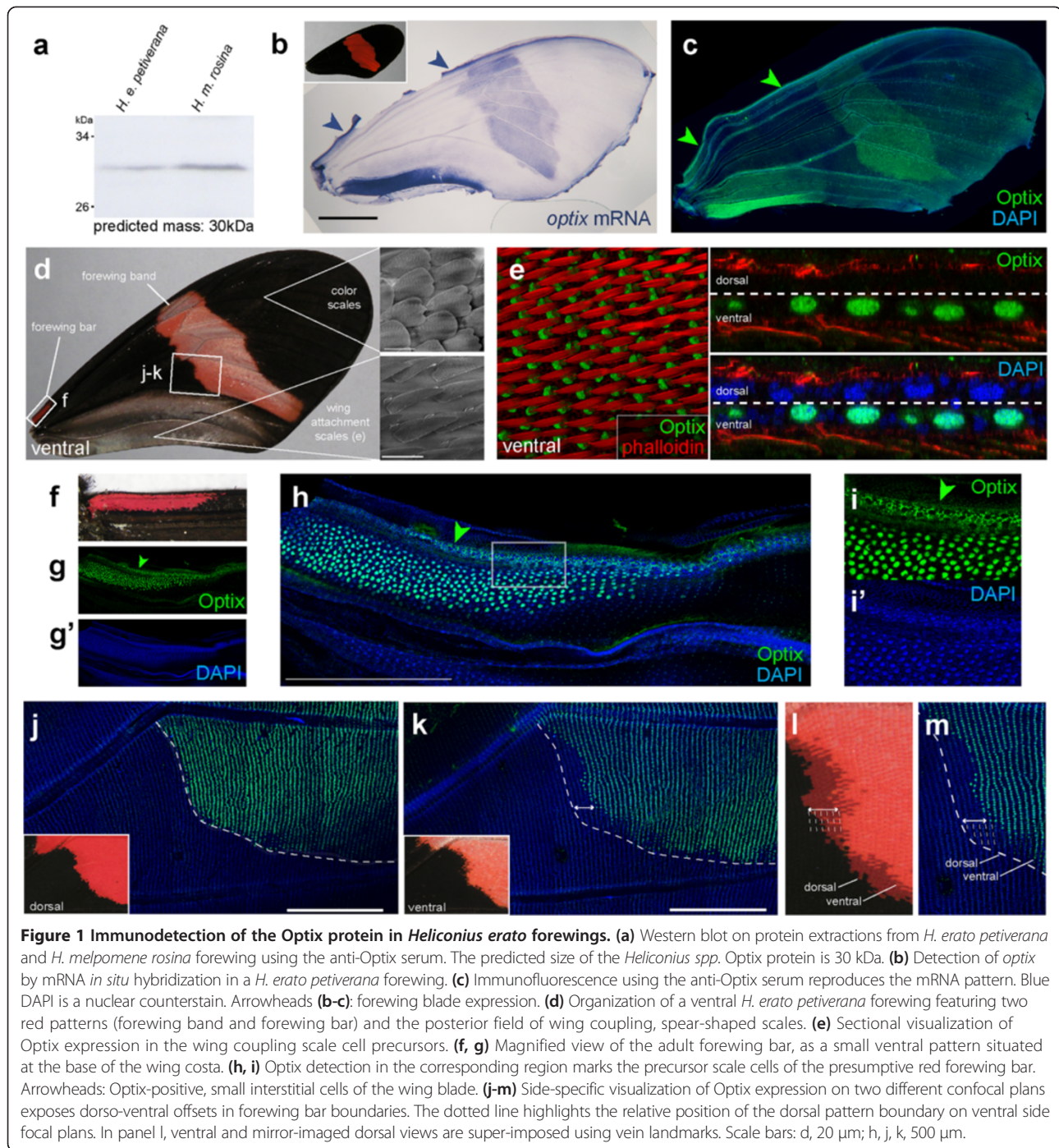
Other methods are described in the Additional file 1: Methods section of this manuscript.

Results

Specific detection of the Optix protein during wing development

A polyclonal antibody serum was generated in a rat immunized with the *Heliconius* Optix protein. The crude serum immunodetected a band of the expected size (30 kDa) in a western blot (Figure 1a), and immunofluorescence in the developing wings of *H. erato petiverana* precisely replicated the *optix* mRNA expression pattern (Figure 1b-c). No signal was observed in *Drosophila melanogaster* and *Danio rerio* embryos (data not shown), suggesting that the polyclonal serum targets epitopes that are Lepidoptera-specific and is also unlikely to detect other lepidopteran SIX family transcription factors. Wing veins and scale sockets produced signals in several negative controls [see Additional file 1: Figure S1], and are thus non-specific signals inherent to our pupal stage immunodetection assays. The scale socket artifactual staining is stage-dependent and does not interfere with the specific detection of Optix at more internal focal planes using confocal imaging, or during whole-wing imaging at lower magnifications using epifluorescence imaging.

Optix was detected during pupal wing development in discrete expression domains that were not associated with color scales. As previously noted [29], Optix marks the large nuclei of cells that differentiate into spear-shaped acute scales (Figure 1d-e) thought to be involved in wing coupling by frictional effect [43]. Optix was also detected in an anterior domain of small interstitial cells that run along the wing blade of the forewing (Figure 1f-i). In terms of color patterns, Optix was associated with presumptive red scales, including in a small ventral red bar at the base of the forewing costa (Figure 1f-h). We confirmed presumptive color scale correspondence with cellular resolution and in a dorsal-ventral specific-manner. For instance, the red median forewing band of *H. erato petiverana* differs on its proximal boundary by up to six scale rows between the ventral and dorsal sides. This difference is reflected by corresponding shifts of Optix immunolocalization (Figure 1j-m). These observations provide further verification at the protein level that *optix* is a selector gene switching on red scale color phenotypes [29,44].



Conserved color pattern-independent roles of Optix across nymphalid butterflies

We compared the expression of Optix during the pupal wing development of 13 butterfly morphs and species of the family Nymphalidae (Figure 2; [see Additional file 1: Figure S2]). In each case, Optix marked the presumptive wing coupling scales on ventral forewings and dorsal hindwings. The shape and size of these fields of

specialized scales are sexually dimorphic in *Heliconius*, with a larger size observed in males compared to females [see Additional file 1: Figure S3; Reference 43]. Corresponding variability in the size and relative positions of Optix positive cell fields were observed; for instance, immunodetection of Optix in *H. hecale fornarina* and a *H. melpomene* hybrid [see Additional file 1: Figure S2] revealed domains of expression extending anteriorly

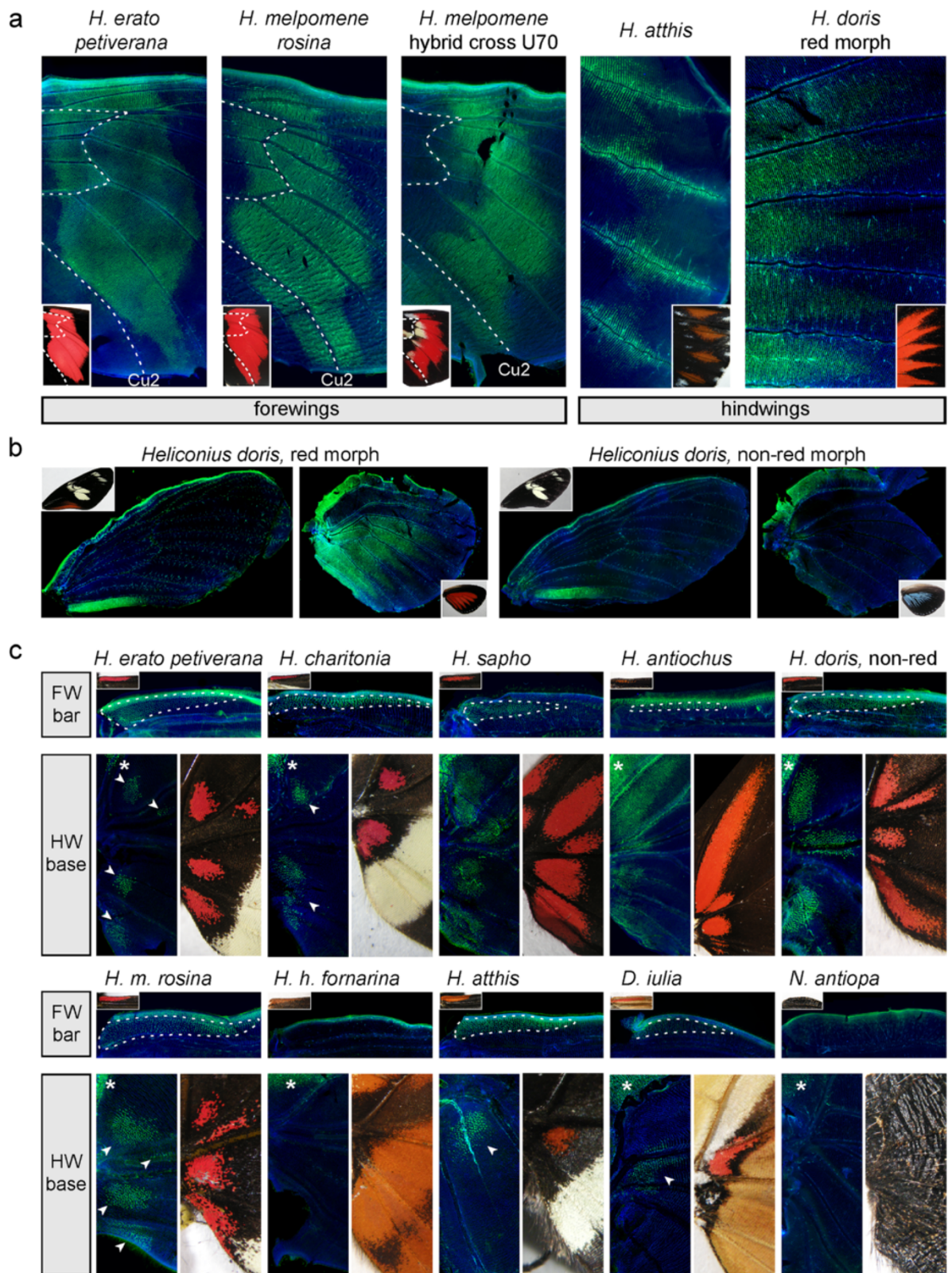


Figure 2 (See legend on next page.)

(See figure on previous page.)

Figure 2 Optix reveals developmental homologies between wing patterns. (a) Optix expression is tightly associated with various *Heliconius* red wing patterns. Notice the interspecific difference in pattern boundaries relative to the Cu2 vein (dotted lines). The hybrid cross U70 is described in Figure 3. (b) Optix expression in the polymorphic *Heliconius doris* butterflies. Insets feature dorsal views of adult patterns. (c) Conserved association of Optix expression with red 'proximal complex' patterns (forewing bar and hindwing basal spots) across Heliconiini. Asterisks: color pattern-independent expression of Optix in dorsal patches of presumptive acute scales. Insets feature ventral views of adult patterns.

on the forewing and posteriorly on the hindwing that are characteristic of male wing coupling patches [see Additional file 1: Figure S3]. In addition, the Optix antigenicity of the forewing wing blade interstitial cells was observed in all tested species. Expression of Optix in the wing blade and wing attachment scale fields replicate previous *optix* mRNA localization results [29], uncover broadly conserved features of butterfly wing pattern development, and provide internal controls for the successful detection of Optix in wing tissues.

Optix reveals homology of red wing patterns among passion-vine butterflies

We replicated the previous observation that Optix expression is associated with the development of red scale patches in *Heliconius* (Figure 2a), and is in large part responsible for convergent evolution of the Central American *H. erato petiverana* and *H. melpomene rosina* co-mimics [29,45]. To further assess wing pattern homologies across the genus *Heliconius* we examined the association of Optix with red coloration to additional species and morphs (Figure 2; [see Additional file 1: Figure S2]), including *Heliconius atthis*, which has remarkably divergent red patterns on the hindwing. One notable species we examined, *Heliconius doris*, is characterized by polymorphic populations where ventral hindwings either display a conspicuous red ray pattern or a more narrow blue or green pattern - discrete variation that is observed within single broods [46]. This variability was present in the stock we assessed (78% red versus 22% non-red individuals; N = 33 emerged butterflies), and was paralleled by Optix immunodetection in a large and rayed ventral patch in 70% (N = 10) of assessed individuals (Figure 2b). Preliminary observations suggest a Mendelian mode of inheritance of *H. doris* wing patterns [46], and our result suggests that allelic variation in *optix* or its upstream regulators is a candidate mechanism for this case of morphological polymorphism.

Optix was consistently detected (with one exception, see below for discussion of *Heliconius hecale fornarina*) in what we term the 'proximal complex', consisting of the red costal forewing bars and of basal hindwing red spots, both situated on ventral wing surfaces (Figures 1j-m and 2c). It is unclear whether such small color elements always have an adaptive function in mimicry, but the clear homology between this set of Optix-positive ventral patterns highlights a conserved

theme of an otherwise multifarious array of *Heliconius* wing patterns. This character has deeper roots in the phylogeny, as we positively identified a homologue of the proximal complex in *Dryas iulia*, which shows Optix-positive scale precursors that mark ventral patches of salmon pink scales. The basal position of *D. iulia* in the Heliconiini clade [see Additional file 1: Figure S4] implies that a color patterning role of Optix preceded the *Heliconius* radiations, consistent with the observation that many species of the Heliconiini clade harbor unambiguous positional homologs of the proximal complex [see Additional file 1: Figure S5]. Similar patterns are not found in heliconiine outgroup lineages, suggesting that the Optix-positive proximal complex is a synapomorphy of the Heliconiini clade of passion-vine butterflies [see Additional file 1: Figure S5].

Optix is spatially regulated by the N locus

In *H. melpomene* an epistatic interaction between *optix* and the modifier locus *N* results in a narrow forewing red band [14,47]. This two-locus system is interesting as it can generate novel patterns by hybridization. For instance, introgression of a *H. melpomene optix* allele into the *cydno* clade generated a chimeric phenotype due to epistasis with *N* in the hybrid species *Heliconius heurippa*, resulting in a distal localization of forewing red [47]. We sought to determine if this activity of *N* is due to an effect on the distribution of the *optix* gene products, or simply to the repression of red pigment synthesis in an otherwise *optix*-positive pattern, as seen in *H. melpomene plesseni* [29]. To distinguish between these two scenarios we used an artificial hybrid zone strategy [31] to generate a composite phenotype analogous to the yellow/red forewing pattern of *H. heurippa* (Figure 3). We found that a genetic factor of Amazonian origin analogous to the N^N allele [14,48] repressed a proximal domain of *optix* expression, resulting in a 'novel' white/red pattern. We thus conclude that *N*, which has not yet been characterized at a molecular level, is a direct or indirect regulator of *optix* expression.

Optix pinpoints regulatory differences between mimetic orange and red patterns

The hindwings of the butterfly *H. hecale fornarina* show a claw-shaped ventral pattern that is reminiscent of a similar pattern observed in *Heliconius cydno galanthus* [49].

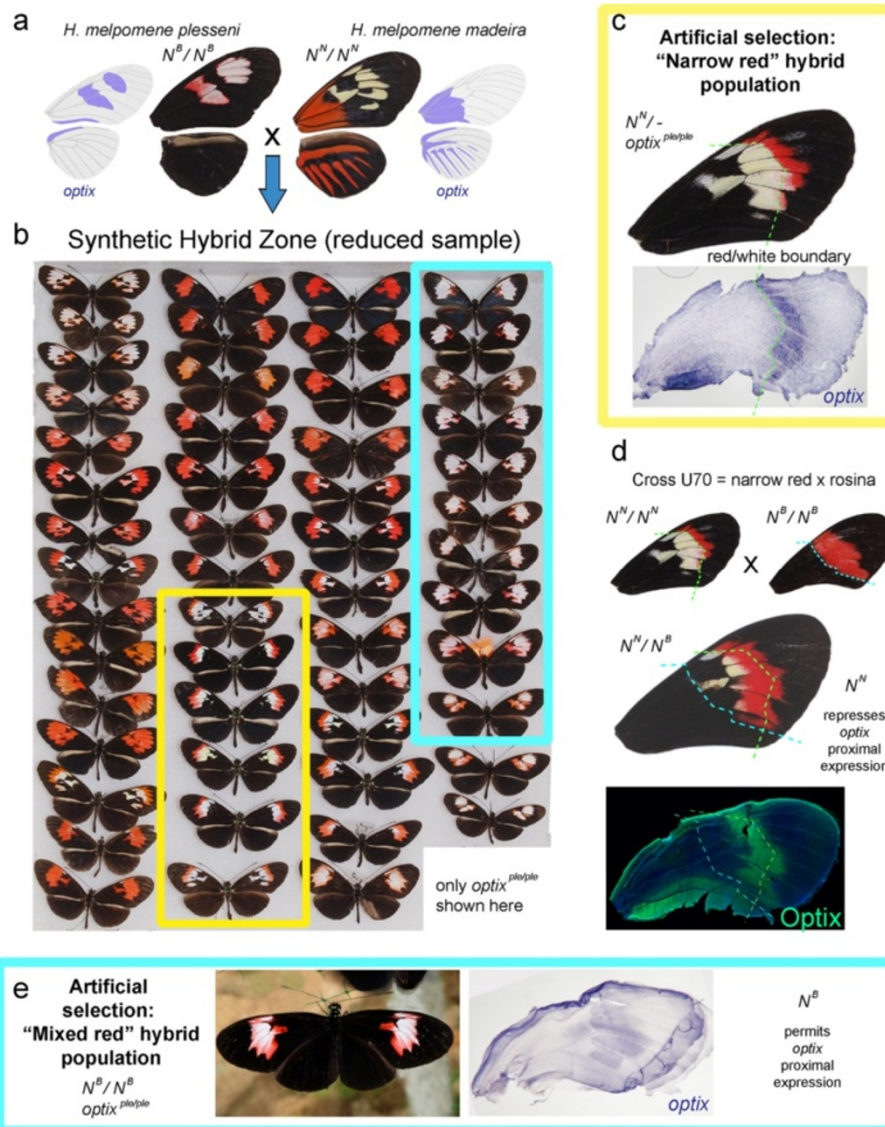


Figure 3 Epistatic regulation of *optix* in hybrid patterns. (a-b) A synthetic hybrid zone was created by interbreeding *Heliconius melpomene plesseni* and *H. melpomene madeira* butterflies in a large greenhouse. Expression of *optix* in founders is reproduced from *in situ* hybridization [29] and the *plesseni* white scales expresses *optix*. The *N* locus that modulates the color composition of the median forewing: non-Amazonian (non-rayed) races such as *plesseni* bear N^B alleles [48], while Amazonian (rayed) races such as *madeira* bear N^N alleles that result in a distal localization of the forewing red band [14,48]. Panel b shows a sample of the phenotypic diversity observed on forewings of this hybrid population. Only butterflies that were homozygous for the *plesseni* *optix* allele are shown here (dominant rayed alleles excluded). Hindwings show no pattern variation in this population and are not shown. **(c)** Artificial selection of butterflies showing a band of distal red (yellow box) generated a stable population called 'Narrow red'. The distal localization of red is explained by N^N alleles that repress proximal expression of *optix*. **(d)** Crossing a 'narrow red' individual with a N^B/N^B butterfly does not recover the full expression of the forewing band red in the F1 progeny, in which *Optix* expression is limited to the distal portion of the white/red boundary. The absence of fully red forewing bands similar to the parental *rosina* pattern in the F1 shows the 'narrow red' parent is N^N -homozygous. **(e)** Artificial selection of butterflies showing a mixture of red and white scales in the forewing band (blue box) generated a stable population called 'Mixed red'. In the presence of N^B alleles derived from *plesseni*, proximal *optix* expression (for instance overlapping with the discal cell) and only delimited by the total shape of the forewing bar.

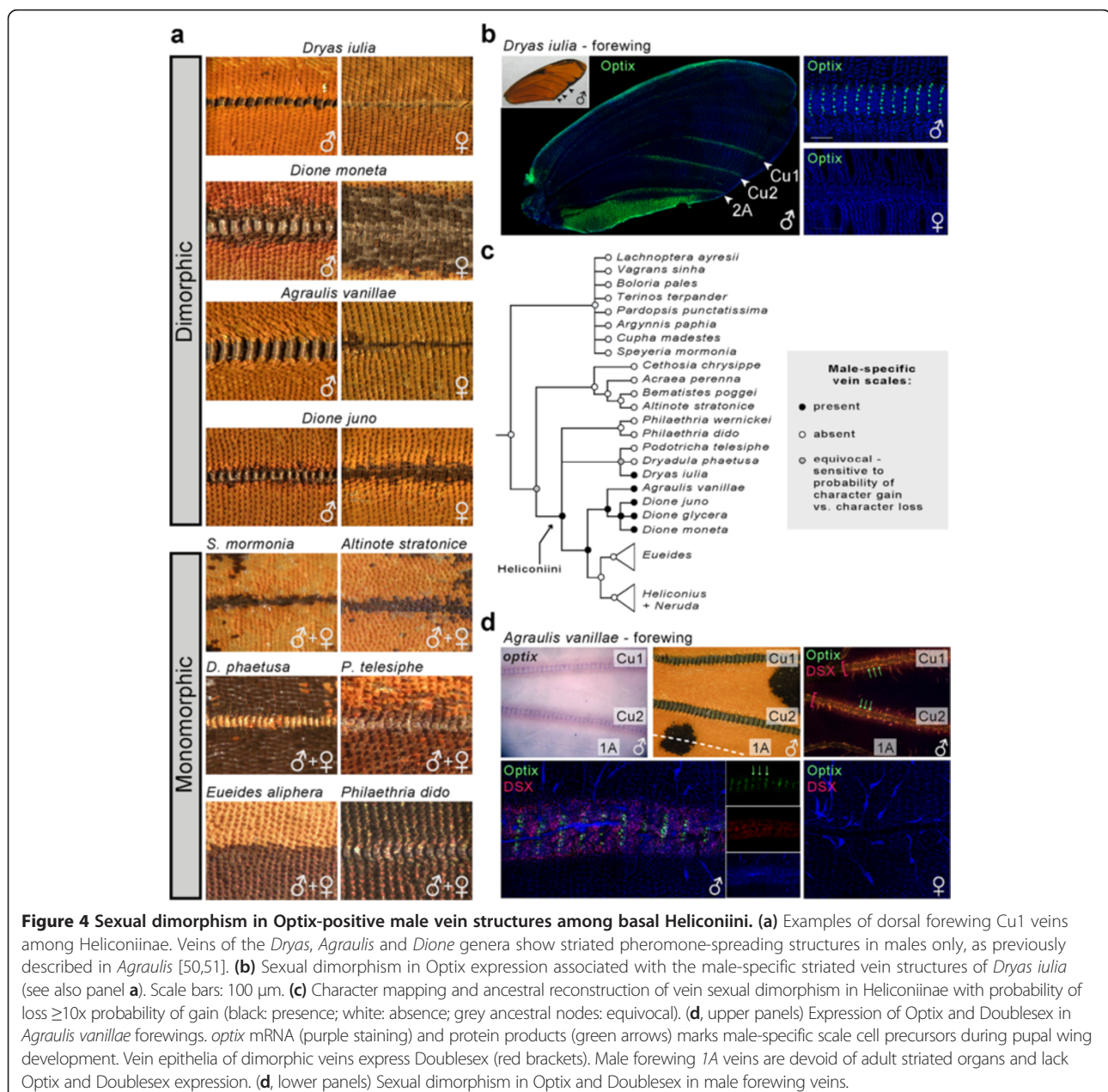
Both butterflies co-occur in a large distribution range that spans southern Mexico, Guatemala, and El Salvador, but differ by the yellow versus white color of their forewing bands. However, spectrometer reflectance measurements performed on the *H. cydno galanthus* and

H. hecale fornarina red hindwing patterns [see Additional file 1: Figure S6] are almost identical and suggest that natural predators are unable to discriminate between these signals. We previously found that the orange-brown claw pattern of *H. cydno* expresses *optix*

[29], and thus wanted to determine whether the *H. hecale fornarina* pattern would reveal a similar expression pattern. Unexpectedly, we found that Optix was neither expressed in the hindwing claw-shaped pattern nor in the short costal forewing bar (Figure 2c). This negative result indicates that *H. hecale fornarina* orange/brown color patterning may follow a divergent process. For example, Optix expression could occur at later developmental stages that could not be assayed by our immunostainings. Whatever the explanation, the color patterning role of Optix in silvaniforms - the tiger-like butterflies similar to *H. hecale* [49] - remains to be determined.

Optix marks presumptive male-specific wing scales in basal Heliconiini

Immunodetection of Optix in *Dryas* marked rows of 6 to 8 scale precursor cells that are dispersed along the Cu1, Cu2, and 2A forewing veins (Figure 4a-b). This expression pattern was observed in male but not in female individuals (N = 5; 3 males, 2 females), suggesting a case of sexual dimorphism. Examination of *Dryas* adult wings revealed the presence of male-specific, striated features on the M3, Cu1, and 2A veins; these features resemble pheromone-spreading structures that were previously described in *Agraulis vanillae* [50,51], another



passion-vine butterfly. Upon closer examination, we noticed that *Optix* was expressed in macronuclear cells (characteristic of scale cell precursors) that cover the dorsal side of the veins, forming rows of 6 to 8 cells that align with extravenous *Optix*-negative scale cell precursors. This relative arrangement was compared to dissections of adult male veins and indicates that *Optix* marks the scale precursors of the male-specific 'scale section-1' [50]. These cells give rise to single rows of 6 to 8 densely packed scales of gold and black color that act as a protective sheath for fragile brush-bearing scales situated underneath [50,51]. This combined scale arrangement is coupled with a pore system and is likely involved in the dissemination of male pheromones; it is also possible that the scale section-1 prevents excessive evaporation of the secreted volatiles [50]. Striated expression of *optix* in this scale system was replicated in *Agraulis*, and we observed similar male-specific structures in the *Dione* genus (Figure 4b). Current tree topology of basal Heliconiini suggests that these *Optix*-positive, sexually dimorphic organs are ancestral in Heliconiini and were lost at least three times during the subsequent radiation of this lineage (Figure 4c).

To gain further insights into the development of sexual dimorphism in the wing veins of these butterflies, we sought to test the expression of *Doublesex*, a transcription factor that has been proposed as a master regulator of sex-specific gene expression and development [52]. An antibody targeting the *Drosophila* *Doublesex* DNA-binding domain was recently developed using a conserved antigen [42]. We hypothesized that this antibody would be likely to cross-react with a butterfly *Doublesex* orthologue, as a BLASTP analysis using the antigenic peptide sequence [42] as a query resulted a single hit with 52 out of 59 conserved residues (88% conservation) against the *Heliconius melpomene* annotated protein set [15]. Remarkably, immunodetection of *Doublesex* in *Agraulis* developing pupal wings specifically marked the dorsal epithelium of the dimorphic veins that include

the *Optix*-positive scale cell precursors (Figure 4c). Only male individuals showed a positive signal among assessed individuals (N = 6, 3 males, 3 females), and all the male veins that are devoid of striated structures - that is, the forewing 1A and all the hindwing veins [50,51] - showed no *Doublesex* signal, revealing a tight association of *Doublesex* expression with sexually dimorphic structures in this system. This result suggests that *Doublesex* activates the development of the male-specific wing scales via its sex-limited expression, and we speculate that *Doublesex* could be a transcriptional co-regulator of *optix* in basal heliconiines.

Optix is expressed in neural progenitors of the optic lobe

It was previously proposed that butterfly wing pigments originate from the redeployment in wings of biosynthetic modules already present in eyes [53]. Indeed, insect eyes express ommochrome pigments that also compose the yellow, orange, and red pigments of *Heliconius* wings [54,55]. Since *optix* is sufficient to induce ectopic eyes that include red pigmentation in flies [56], it was proposed that the co-option of a simplified *optix* eye regulatory module could have taken place in wings, therefore explaining the regulation of red ommochrome pigmentation by *optix* in *Heliconius* [57]. We assessed a variant of this 'eye-to-wing' co-option hypothesis by looking at *optix* expression in pupal eyes at the onset of ommochrome pigment deposition, as well as in adult retinas, where red pigment granules occur [58]. While *optix* is transcribed in wings at the pupal stage we examined [29,59], we did not detect *Optix* immunoreactivity in pupal or adult retinas of various *Heliconius* species (Figure 5). Interestingly, however, *Optix* was expressed at the pre-ommochrome stage in two single layers of neural progenitors that bordered the inner rim of the lamina and the outer rim of the medulla (Figure 5a). These neuroepithelia are the first integrators of visual information, and it will be interesting to further use *Optix* as a marker of

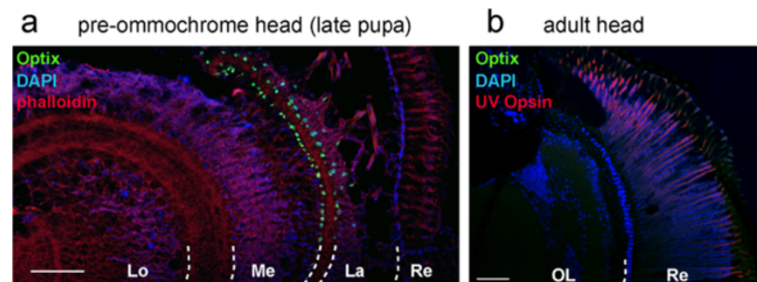


Figure 5 *Optix* is transiently expressed in a small population of optic lobe neurons. (a) Expression of *Optix* in *Heliconius* optic lobes at the pre-ommochrome stage. Similar results were obtained for *H. doris* and *H. sapho*. (b) Lack of *Optix* expression in optic lobes and retina at the adult stage. Immunodetection of the photoreceptors UVRh1/2 is shown here in ommatidia as a positive control. This negative result was replicated in *H. melpomene*, *H. cydno*, and *H. doris*. La, lamina; Lo, lobula; Me, medulla; Re, retina. OL, optic lobe. Scale bars: 100 μ m.

butterfly neuronal patterning and determine its role in visual processing.

Discussion

The evolutionary origin of red wing patterns in *Heliconius* is unclear, as these elements seem divergent from the common pattern scheme shared by other butterflies [37]. Here, we clarify this issue by (a) arguing that Optix-positive patterns are a relatively recent novelty that seemingly appeared by co-option of Optix expression in color scale cell precursors, and (b) disentangling different genetic modes of pattern diversification that followed the original co-option event.

Optix color patterns are an evolutionary novelty of passion-vine butterflies

The co-option of a gene into a new context constitutes an unambiguous example of evolutionary novelty at the molecular level [60]. In particular, the re-deployment of developmental genes has been proposed to underlie the origin of novel traits such as butterfly wing patterns [10,61]. We observed Optix in the precursors of three types of scales: ancestral spear-shaped wing coupling scales [29]; specialized, male-specific, gold and black scales that are proposed to protect pheromone-spreading structures in basal Heliconiini [50]; and finally, the colored scales associated with pink (*Dryas iulia*) or red (*Heliconius spp.*) pigmentation across Heliconiini. We propose that each of these three Optix-associated scale phenotypes forms a distinct evolutionary character due to the divergent scale morphologies they underlie. Importantly, the recruitment of Optix expression into colored scale cell precursors of the 'proximal complex' is a synapomorphy of neotropical passion-vine butterflies (Heliconiini) and was detected in the basal genus *Dryas*, thus marking an innovation event associated with the Heliconiini radiation (Figure 6). In *H. erato* and *H. melpomene*, *optix* expression may activate the *cinnabar* and *ebony* pigment synthesis genes, as their transcripts show a strong spatial correlation with Optix-positive red pigmentation [54,55,59,62]. It will be interesting to further confirm the regulatory targets of Optix and to determine how many steps were necessary to achieve its color patterning role. As we were unable to validate the hypothesis of an eye-to-wing co-option event re-wiring an ancient pigmentation module [57], the mechanism underlying the recruitment of Optix into a color determination module remains to be elucidated.

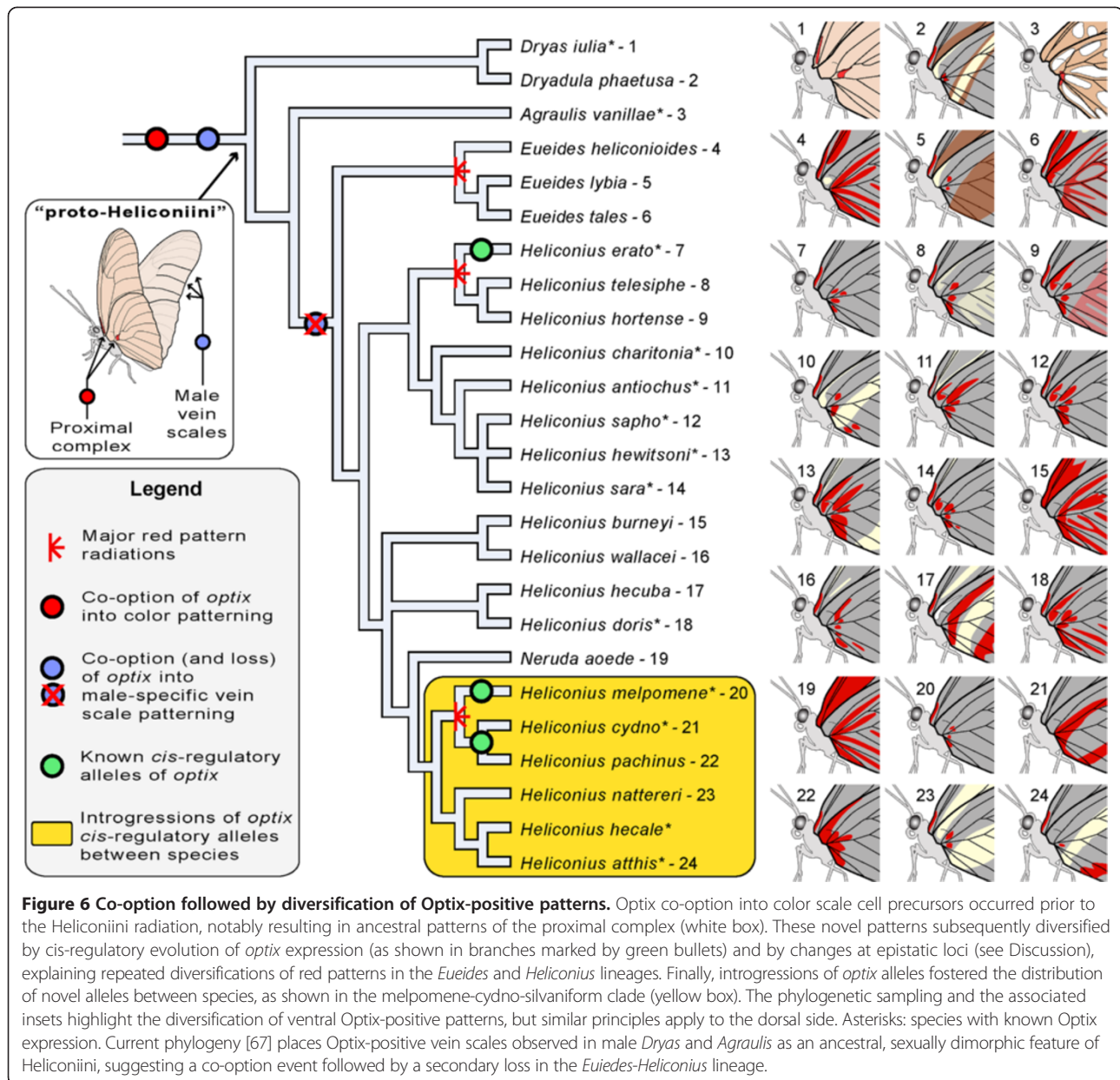
In parallel with its recently evolved color patterning role, Optix also acquired a novel role in the specification of male-specific striated scale structures on the forewing veins of *Agraulis* butterflies as well as in the basal genus *Dryas*, suggesting a co-option event prior to the Heliconiini radiation (Figure 6). This implies that

Optix has been repeatedly recruited in the specification of derived scale identities, highlighting a possible propensity of the *optix* gene to be recruited during the evolution of specialized scale types.

Four modes of red pattern diversification in *Heliconius*

Optix expression implies that all the red patterns assessed in the *Heliconius* genus are variations of a conserved core process linking Optix expression in scale cell precursors to a narrow range of scale phenotypes (Figure 6). Under the conceptual framework of developmental homology [63] Optix scale cell precursor expression provides a character-defining mechanism (that is, the presence of red scales) that pinpoints an evolutionary novelty, whereas modulation of Optix spatial localization underlies the 'character states' of this trait and is a secondary effect of diversification. We summarize here our knowledge of the different modes of evolution of red patterns with a focus on *optix* regulation:

1. *cis*-regulatory evolution of *optix* has played a predominant role in the tinkering of novel red patterns. At least eight *optix* alleles with distinct pattern-switching effects have been identified by genetic mapping: four alleles in the *H. erato/H. himera* clade [27], two alleles in *H. melpomene* [25], and two alleles in the *H. cydno/Heliconius pacheus* sister species [26]. It is proposed that this allelic diversity results from the cumulative effect of many tightly linked *cis*-regulatory sequence variations organized in complex haplotypes and possibly compartmentalized into several pattern-specific *cis*-regulatory modules [45]. Following the observation that the genetic basis of phenotypic variation is highly repeatable [33], and that *optix* regulation maps to a large adjacent intergenic region that could reflect a potential for complex *cis*-regulatory structure, we extrapolate that direct *cis*-regulatory evolution of *optix* may extend to many cases of red pattern heterotopy (spatial shifts) in *Heliconius*.
2. While the *optix* locus tends to explain presence/absence polymorphism of a given pattern element (for example, the forewing band), pattern shape variation can sometimes be explained by epistatic effects with unlinked loci. Several examples have been characterized at various levels. For instance, the early expression of *WntA* during larval development determines the shape of the red forewing band [27,64]. We also show here that *N* spatially regulates Optix pattern shapes in *H. melpomene*.
3. Adaptive *optix* alleles have been shown to introgress across geographic barriers and to create taxonomic mosaics of pattern diversity, where a given pattern of single origin is scattered across the lineage



phylogeny [18]. More surprisingly, lateral transfer by hybridization of wing patterns can occur beyond the species boundary [31]. Directly quoting the original report by Gilbert (2003), *virtually all departures from the common pattern themes within the cydney clade (for example, H. heurippa, H. timareta, H. pacheus, H. cydney weymeri form gustavi) can be explained by introgression of toolbox elements from H. melpomene*. *Optix* appears to be a core component of this toolbox, and recent genetic data has since confirmed that cross-species introgressions of *optix* alleles drive multiple examples of mimetic convergence in the ‘melpomene-cydney-silvaniform’ clade [15,30,32,65]. Introgressions do not directly

cause the evolution of novel patterning alleles, but propagate adaptive variants across gene pools, and hybrids can display effects of novelty due to epistatic effects between wing pattern loci, as seen in *H. heurippa* [47,65]. Introgressions may be a major but previously unsuspected cause of phenotypic disparity across the animal kingdom, and *Heliconius optix* alleles emerge as a model system of choice to study this phenomenon [66].

- In addition to the effects of loci that act upstream of *optix* and affect its localization, modifier loci acting downstream of *optix* may modulate its color output. A likely example occurs in *H. melpomene plesseni*, where an unidentified mechanism converts *optix*-

positive scales into a highly-UV reflective scale field of white appearance [29,48]. This epistatic factor of *plesseni* origin is visible in an artificial 'Mixed white' hybrid population, where an unbroken forewing patch of white and red scales expresses *optix* (Figure 3e).

In summary, 1) cis-regulatory evolution of *optix* and epistatic interactions with upstream trans-regulatory factors affect the localization and shape of red patterns; 2) introgressions of *optix* variants maximize the phylogenetic distribution of pre-existing patterns, sometimes resulting in novel, hybrid patterns due to epistatic effects; and 3) unidentified modifier genes can modulate the color output of *Optix*-positive scale cell precursors. Similar principles may apply to *Optix*-independent patterns.

Conclusions

Multiple lines of evidence suggest that the regulatory evolution of the *optix* locus drives morphological radiations in the wings of *H. melpomene* and *H. erato* [15,18,30,45]. Regulatory evolution of this locus may also have a direct influence on species richness itself due to assortative mating between butterflies displaying identical red wing patterns [35,36]. In the light of the present data, we infer that the evolution of *Optix* color patterning roles sparked the diversification of red wing patterns within the Heliconiini tribe, notably among the spectacular mimetic radiations observed in the genera *Heliconius* and *Eueides*. Multiple recruitments of *Optix* expression into the differentiation of derived wing scales - the red color scale cells, as well as the male-specific vein structures of basal heliconiines - provide compelling examples of developmental innovation, and the genetic co-option event associated to coloration was followed by a rapid radiation in wing patterns. By opening paths to unexplored valleys of the morphospace, gene co-option events may be a general mechanism explaining sudden bursts of phenotypic change.

Additional file

Additional file 1: Martin_et_al-Supplementary.pdf. Supplementary Materials including Supplementary Methods, Supplementary Table, Supplementary Figures 1–6 and Supplementary References.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AM carried out wing development work, analyzed the data and drafted the manuscript. KJM carried out eye and brain immunostainings. NHP supervised antibody production. ADB provided live *Heliconius* pupae and carried out the spectrophotometer measurements. LEG performed the hybrid *Heliconius* crosses and contributed live *Heliconius* pupae. RDR supervised the project and helped to draft the manuscript. All authors read and approved the final manuscript.

Acknowledgements

We are grateful to Angela Kaczmarczyk for helping with recombinant protein purification, Carmen Robinett for sharing the anti-Doublesex antibody, and Theresa Freiburger for butterfly husbandry at UT Austin. Thomas Schilling provided continuous support for this research and we also thank the members of his laboratory for technical assistance and fruitful discussions. This work was supported by the National Science Foundation grant IOS-1052541 to RDR.

Author details

¹Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853, USA. ²Department of Ecology and Evolutionary Biology, University of California, Irvine, CA 92697-2525, USA. ³Department of Molecular and Cell Biology, University of California, Berkeley, CA 94720-3140, USA. ⁴Department of Integrative Biology, University of Texas, Austin, TX 78712, USA.

Received: 28 September 2013 Accepted: 27 November 2013

Published: 5 February 2014

References

1. Simpson GG: *The Major Features of Evolution*. New York: Columbia University Press; 1953.
2. Schluter D: *The Ecology of Adaptive Radiation*. New York: Oxford University Press; 2000.
3. Darwin C: *Voyage of the Beagle*. New York: Collier; 1909.
4. Wheat CW, Vogel H, Wittstock U, Braby MF, Underwood D, Mitchell-Olds T: **The genetic basis of a plant–insect coevolutionary key innovation.** *Proc Natl Acad Sci* 2007, **104**:20427–20431.
5. Blount ZD, Barrick JE, Davidson CJ, Lenski RE: **Genomic analysis of a key innovation in an experimental *Escherichia coli* population.** *Nature* 2012, **489**:513–518.
6. Meyer JR, Dobias DT, Weitz JS, Barrick JE, Quick RT, Lenski RE: **Repeatability and contingency in the evolution of a key innovation in phage Lambda.** *Science* 2012, **335**:428–432.
7. Carlson BA, Hasan SM, Hollmann M, Miller DB, Harmon LJ, Arnegard ME: **Brain evolution triggers increased diversification of electric fishes.** *Science* 2011, **332**:583–586.
8. Arnoult L, Su KFY, Manoel D, Minervino C, Magriña J, Gompel N, Prud'homme B: **Emergence and diversification of fly pigmentation through evolution of a gene regulatory module.** *Science* 2013, **339**:1423–1426.
9. Near TJ, Dornburg A, Kuhn KL, Eastman JT, Pennington JN, Patarnello T, Zane L, Fernández DA, Jones CD: **Ancient climate change, antifreeze, and the evolutionary diversification of Antarctic fishes.** *Proc Natl Acad Sci* 2012, **109**:3434–3439.
10. Monteiro A, Podlaha O: **Wings, horns, and butterfly eyespots: how do complex traits evolve?** *PLoS Biol* 2009, **7**:e1000037.
11. Moczek AP: **On the origins of novelty in development and evolution.** *BioEssays* 2008, **30**:432–447.
12. Kopp A: **Metamodels and phylogenetic replication: a systematic approach to the evolution of developmental pathways.** *Evolution* 2009, **63**:2771–2789.
13. Joron M, Jiggins CD, Papanicolaou A, McMillan WO: **Heliconius wing patterns: an evo-devo model for understanding phenotypic diversity.** *Heredity* 2006, **97**:157–167.
14. Sheppard PM, Turner JRG, Brown KS, Benson WW, Singer MC: **Genetics and the evolution of Müllerian mimicry in Heliconius butterflies.** *Philos Trans R Soc Lond Ser B-Biol Sci* 1985, **308**:433.
15. Heliconius Genome Consortium: **Butterfly genome reveals promiscuous exchange of mimicry adaptations among species.** *Nature* 2012, **487**:94–98.
16. Brown KS: *Ecologia Geográfica e Evolução nas Florestas Neotropicais*. Campinas, Brazil: Universidade Estadual de Campinas; 1979.
17. Bates HW: **Contributions to an insect fauna of the Amazon valley (Lepidoptera: Heliconiidae).** *Trans Linn Soc Lond*. 1862, **23**:495–566.
18. Hines HM, COUNTERMAN BA, PAPA R, CARDOSO MZ, LINARES M, MALLET J, REED RD, JIGGINS CD, KRONFORST MR, McMillan WO: **Wing patterning gene redefines the mimetic history of Heliconius butterflies.** *Proc Natl Acad Sci U S A* 2011, **108**:19666–19671.
19. Mallet J, Gilbert LE: **Why are there so many mimicry rings? Correlations between habitat, behaviour and mimicry in Heliconius butterflies.** *Biol J Linn Soc* 1995, **55**:159–180.
20. Lamas G, Callaghan C, Casagrande MM, Mielke O, Pycrz T, Robbins R, Viloria A: **Hesperioidea - Papilionoidea.** In *Atlas Neotropical Lepidoptera Checklist Part 4A*.

- Edited by Heppner JB. Gainesville, Florida: Association for Tropical Lepidoptera/Scientific Publishers; 2004.
21. Gilbert LE: **Biodiversity of a Central American Heliconius community: pattern, process, and problems.** Chap. 18. In *Plant-animal Interactions: Evolution Ecology in Tropical and Temperate Regions*. Edited by Price PW, Lewinsohn TM, Fernandes TW, Benson WW. New York: John Wiley and Sons; 1991:403–427.
 22. Spencer KC: **Chemical mediation of coevolution in the Passiflora-Heliconius interaction.** In *Chemical mediation of coevolution*. Edited by Spencer KC. London: Academic; 1988:167–240.
 23. Benson WW, Brown KS, Gilbert LE: **Coevolution of plants and herbivores: passion flower butterflies.** *Evolution*. 1975, **29**:659–680.
 24. Joron M, Iwasa Y: **The evolution of a Müllerian mimic in a spatially distributed community.** *J Theor Biol* 2005, **237**:87–103.
 25. Baxter S, Papa R, Chamberlain N, Humphray S, French-Constant R, McMillan WO: **Parallel evolution in the genetic basis of Mullerian mimicry in Heliconius butterflies.** *Genetics* 2008, **180**:1567–1577.
 26. Chamberlain NL, Hill RI, Baxter SW, Jiggins CD, Kronforst MR: **Comparative population genetics of a mimicry locus among hybridizing Heliconius butterfly species.** *Heredity* 2011, **107**:200–204.
 27. Papa R, Kapan DD, Counterman BA, Maldonado K, Lindstrom DP, Reed RD, Nijhout HF, Hrbek T, McMillan WO: **Multi-allelic major effect genes interact with minor effect QTLs to control adaptive color pattern variation in Heliconius erato.** *PLoS ONE* 2013, **8**:e57033.
 28. Papa R, Martin A, Reed RD: **Genomic hotspots of adaptation in butterfly wing pattern evolution.** *Curr Opin Genet Dev* 2008, **18**:559–564.
 29. Reed RD, Papa R, Martin A, Hines HM, Counterman BA, Pardo-Diaz C, Jiggins CD, Chamberlain NL, Kronforst MR, Chen R, Halder G, Nijhout HF, McMillan WO: **optix drives the repeated convergent evolution of butterfly wing pattern mimicry.** *Science* 2011, **333**:1137–1141.
 30. Pardo-Diaz C, Salazar C, Baxter SW, Merot C, Figueiredo-Ready W, Joron M, McMillan WO, Jiggins CD: **Adaptive introgression across species boundaries in Heliconius butterflies.** *PLoS Genet* 2012, **8**:e1002752.
 31. Gilbert LE: **Adaptive novelty through introgression in Heliconius wing patterns: evidence for shared genetic “tool box” from synthetic hybrid zones and a theory of diversification.** In *Butterflies: Ecology and Evolution Taking Flight*. Edited by Boggs CL, Watt WB, Ehrlich PR. Chicago: University of Chicago Press; 2003:281–318.
 32. Smith J, Kronforst MR: **Do Heliconius butterfly species exchange mimicry alleles?** *Biol Lett* 2013, **9**:20130503.
 33. Martin A, Orgogozo V: **The loci of repeated evolution: a catalog of genetic hotspots of phenotypic variation.** *Evolution* 2013, **67**:1235–1250.
 34. Melo MC, Salazar C, Jiggins CD, Linares M: **Assortative mating preferences among hybrids offers a route to hybrid speciation.** *Evolution* 2009, **63**:1660–1665.
 35. Jiggins CD, Naisbit RE, Coe RL, Mallet J: **Reproductive isolation caused by colour pattern mimicry.** *Nature* 2001, **411**:302–305.
 36. Merrill RM, Van Schooten B, Scott JA, Jiggins CD: **Pervasive genetic associations between traits causing reproductive isolation in Heliconius butterflies.** *Proc R Soc B Biol Sci* 2011, **278**:511–518.
 37. Nijhout HF, Wray GA: **Homologies in the color patterns of the genus Heliconius (Lepidoptera, Nymphalidae).** *Biol J Linn Soc*. 1988, **33**:345–365.
 38. Reed RD, Serfas MS: **Butterfly wing pattern evolution is associated with changes in a Notch/Distal-less temporal pattern formation process.** *Curr Biol* 2004, **14**:1159–1166.
 39. Martin A, Reed RD: **wingless and aristaless2 define a developmental ground plan for moth and butterfly wing pattern evolution.** *Mol Biol Evol* 2010, **27**:2864–2878.
 40. Otaki JM: **Color pattern analysis of nymphalid butterfly wings: revision of the nymphalid groundplan.** *Zoolog Sci* 2012, **29**:568–576.
 41. Lampel J, Briscoe A, Wasserthal L: **Expression of UV-, blue-, long-wavelength-sensitive opsins and melanin in extraretinal photoreceptors of the optic lobes of hawkmoths.** *Cell Tissue Res* 2005, **321**:443–458.
 42. Mellert DJ, Robinett CC, Baker BS: **doublesex functions early and late in gustatory sense organ development.** *PLoS ONE* 2012, **7**:e51489.
 43. Bahi Aymone AC, Valente Gaisky VLS, Mellender de Araújo A: **Ultrastructure and morphogenesis of the wing scales in Heliconius erato phyllis (Lepidoptera: Nymphalidae): What silvery/brownish surfaces can tell us about the development of color patterning?** *Arthropod Struct Dev* 2013, **42**:349–359.
 44. Mann RS, Carroll SB: **Molecular mechanisms of selector gene function and evolution.** *Curr Opin Genet Dev* 2002, **12**:592–600.
 45. Supple MA, Hines HM, Dasmahapatra KK, Lewis JJ, Nielsen DM, Lavoie C, Ray DA, Salazar C, McMillan WO, Counterman BA: **Genomic architecture of adaptive color pattern divergence and convergence in Heliconius butterflies.** *Genome Res* 2013, **23**:1248–1257.
 46. Sheppard PM: **Some genetic studies of müllerian mimics in butterflies of the Heliconius genus.** *Zoologica* 1963, **48**:145–154.
 47. Mavárez J, Salazar CA, Bermingham E, Salcedo C, Jiggins CD, Linares M: **Speciation by hybridization in Heliconius butterflies.** *Nature* 2006, **441**:868–871.
 48. Salazar PA: *Hybridization and the genetics of wing colour-pattern diversity in Heliconius butterflies*. Cambridge, UK: Cambridge University; 2012.
 49. Brown KS: **An illustrated key to the silvaniform Heliconius (Lepidoptera: Nymphalidae) with descriptions of new subspecies.** *Trans Am Entomol Soc* 1976, **102**:373–484.
 50. Rauser CL, Rutowski RL: **Male-specific scales on the wings of the Gulf Fritillary Butterfly, Agraulis vanillae (Lepidoptera: Nymphalidae): pheromone-disseminating structures?** *J Lepidopterists Soc* 2003, **57**:279–283.
 51. Müller F: **The scent-scales of the male of Dione vanillae.** *Kosmos* 1877, **2**:38–41. Translation in: Longstaff, G. B. 1912. *Butterfly Hunting in Many Lands*. Longmans, Green, and Co., London.
 52. Kopp A: **Dmr genes in the development and evolution of sexual dimorphism.** *Trends Genet* 2012, **28**:175–184.
 53. Reed RD, Nagy LM: **Evolutionary redeployment of a biosynthetic module: expression of eye pigment genes vermilion, cinnabar, and white in butterfly wing development.** *Evol Dev* 2005, **7**:301–311.
 54. Ferguson LC, Jiggins CD: **Shared and divergent expression domains on mimetic Heliconius wings.** *Evol Dev* 2009, **11**:498–512.
 55. Reed RD, McMillan WO, Nagy LM: **Gene expression underlying adaptive variation in Heliconius wing patterns: non-modular regulation of overlapping cinnabar and vermilion prepatterns.** *Proc R Soc B Biol Sci* 2008, **275**:37–46.
 56. Seimiya M, Gehring WJ: **The Drosophila homeobox gene optix is capable of inducing ectopic eyes by an eyeless-independent mechanism.** *Development* 2000, **127**:1879–1886.
 57. Monteiro A: **Gene regulatory networks reused to build novel traits: co-option of an eye-related gene regulatory network in eye-like organs and red wing patches on insect wings is suggested by optix expression.** *BioEssays* 2012, **34**:181–186.
 58. Zaccardi G, Kelber A, Sison-Mangus MP, Briscoe AD: **Color discrimination in the red range with only one long-wavelength sensitive opsin.** *J Exp Biol* 2006, **209**:1944–1955.
 59. Hines HM, Papa R, Ruiz M, Papanicolaou A, Wang C, Nijhout HF, McMillan WO, Reed RD: **Transcriptome analysis reveals novel patterning and pigmentation genes underlying Heliconius butterfly wing pattern variation.** *BMC Genomics* 2012, **13**:288.
 60. Hall BK, Kerney R: **Levels of biological organization and the origin of novelty.** *J Exp Zool B Mol Dev Evol* 2012, **318**:428–437.
 61. Saenko SV, French V, Brakefield PM, Beldade P: **Conserved developmental processes and the formation of evolutionary novelties: examples from butterfly wings.** *Philos Trans R Soc B Biol Sci* 2008, **363**:1549–1556.
 62. Ferguson LC, Maroja L, Jiggins C: **Convergent, modular expression of ebony and tan in the mimetic wing patterns of Heliconius butterflies.** *Dev Genes Evol* 2011, **221**:297–308.
 63. Wagner GP: **The developmental genetics of homology.** *Nat Rev Genet* 2007 Jun, **8**:473–479.
 64. Martin A, Papa R, Nadeau NJ, Hill RI, Counterman BA, Halder G, Jiggins CD, Kronforst MR, Long AD, McMillan WO, Reed RD: **Diversification of complex butterfly wing patterns by repeated regulatory evolution of a Wnt ligand.** *Proc Natl Acad Sci* 2012, **109**:12632–12637.
 65. Salazar C, Baxter SW, Pardo-Diaz C, Wu G, SurrIDGE A, Linares M, Bermingham E, Jiggins CD: **Genetic evidence for hybrid trait speciation in Heliconius butterflies.** *PLoS Genet* 2010, **6**:e1000930.
 66. Hedrick PW: **Adaptive introgression in animals: examples and comparison to new mutation and standing variation as sources of adaptive variation.** *Mol Biol Evol* 2013, **22**:4606–4618.
 67. Yuan F, Bernard GD, Le J, Briscoe AD: **Contrasting modes of evolution of the visual pigments in Heliconius butterflies.** *Mol Biol Evol* 2010, **27**:2392–2405.

doi:10.1186/2041-9139-5-7

Cite this article as: Martin et al.: Multiple recent co-options of Optix associated with novel traits in adaptive butterfly wing radiations. *EvoDevo* 2014 **5**:7.